

Rapid DNA Maturity Assessment



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Integration of the extraction, amplification, separation, and detection processes for forensic DNA typing is a challenging goal. Several parallel efforts have been made to integrate the forensic workflow and utilize a simple swab in, answer out process within a single platform [1-3]. Of the multiple efforts, two platforms were tested as a part of a rapid DNA maturity assessment in 2014. The assessment was conducted with sets of blinded single-source reference samples to gauge the typing success of the current rapid DNA typing technology. Samples were provided to participants for testing on the individual rapid platforms, and data was returned to the National Institute of Standards and Technology (NIST) for review and analysis. Both automated and manual review of the data sets were conducted to assess the success of typing the CODIS core loci. Genotyping profiles from the multiple platforms, participants, and STR typing chemistry was combined into a single analysis to assess the current maturity of Rapid DNA technology. The presented results will focus on genotyping success rate, peak height ratios, and stutter artifacts across two platforms and multiple STR kit chemistries.

What is Rapid DNA Typing?

Rapid DNA, or Rapid DNA Analysis, describes the fully automated (hands) free) process of developing a CODIS Core STR profile from a reference sample buccal swab in less than 2 hours. The "swab in – profile out" process consists of automated extraction, amplification, separation, detection and allele calling without human intervention. https://www.fbi.gov/about-us/lab/biometric-analysis/codis/rapid-dna-addendum-to-gas-final-effective-12-1-2014

Rapid DNA Analysis vs. Modified Rapid DNA Analysis

Integrated Rapid DNA Typing Devices Tested

As part of the Maturity Assessment





Rapid DNA analysis describes the fully automated (hands-free) process of developing a CODIS Core STR profile from a known reference sample. The "swab in – profile out" process consists of automated extraction, amplification, separation, detection and allele calling without human intervention.

Modified Rapid DNA analysis describes the automated (hands-free) process of developing a CODIS Core STR profile from a known reference sample. This "swab in – profile out" process consists of automated extraction, amplification, separation, and detection without human intervention but requires manual interpretation and technical review.

https://www.fbi.gov/about-us/lab/biometric-analysis/codis/rapid-dna-addendum-to-gas-final-effective-12-1-2014

ANDE (NetBio)

- Electrophoresis takes place on chip
- Kit = one biochipset

Stored at room temperature

Shelf life \approx 6 months

- RFID swabs tagged for sample tracking

PowerPlex 16 loci ≈86 min runtime (5 samples)

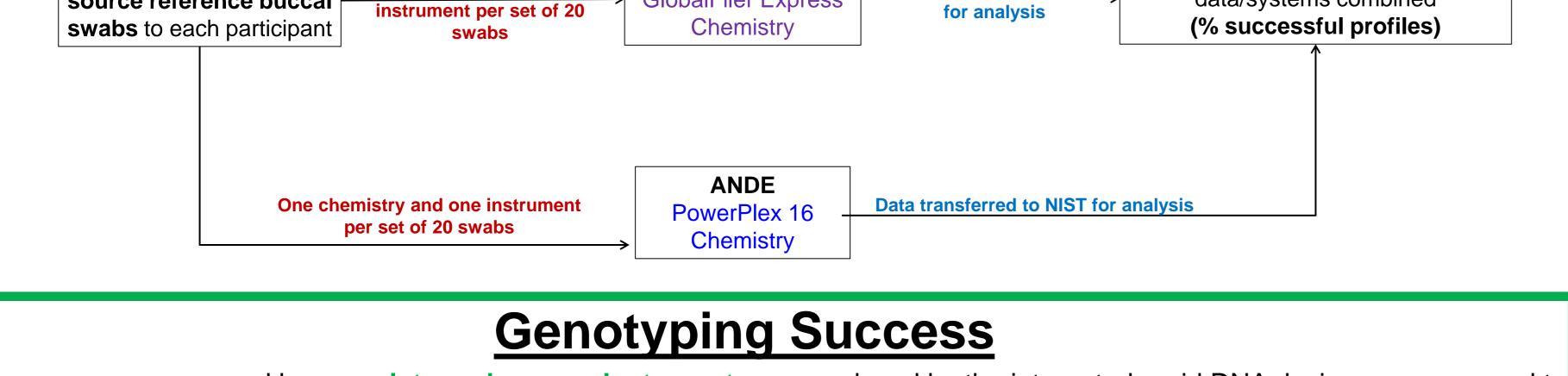
RapidHIT 200 (IntegenX)

- Electrophoresis takes place on an 8 capillary array
- Kit = 4 separate components Stored between room temp and 4°C Shelf life \approx 6 months at 4°C
- Cotton swabs

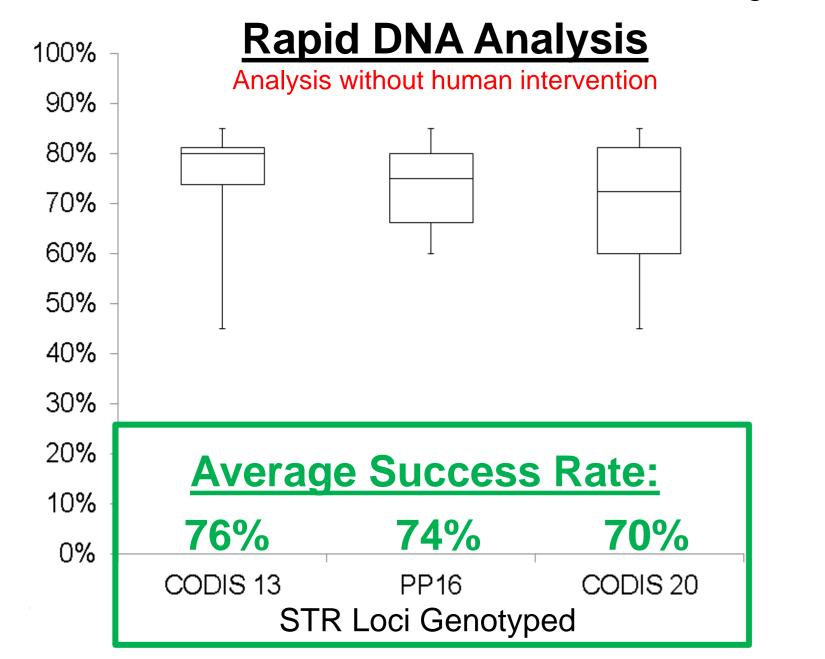
GlobalFiler Express loci PowerPlex 16 loci ≈90 min runtime ≈116 min runtime (5 samples) (1-7 samples)

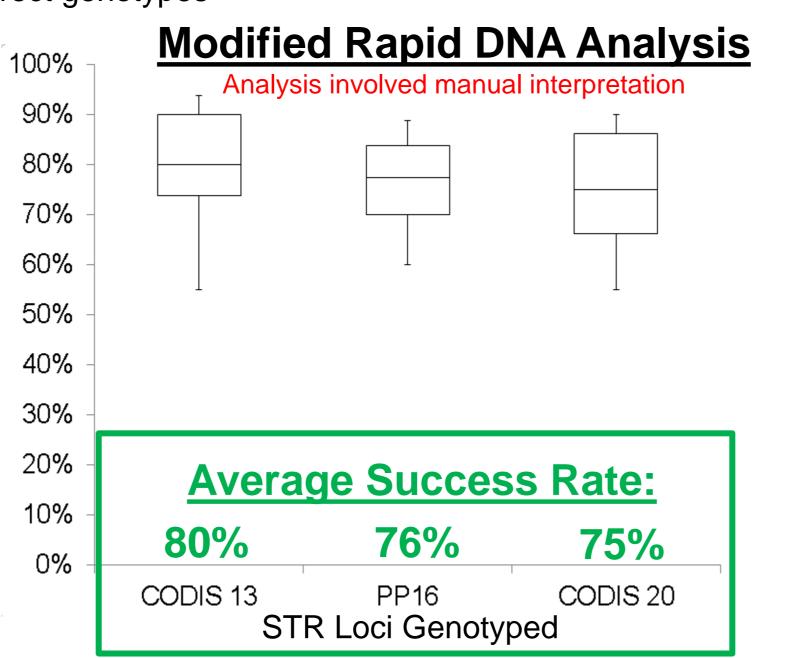
2014 NIST Rapid DNA Maturity Assessment

The purpose of the 2014 NIST Rapid DNA Maturity Assessment was to assess the current status of rapid DNA typing technology for Participating Independent Instrument **Total Samples** Chemistry the CODIS core loci in support of lab and future external (non-lab-based) Rapid DNA instrument implementation. Laboratories (7) Instruments (11) Platforms (2) **Tested (280)** Only integrated (swab in – allele detection) instruments capable of genotyping the core CODIS 13 STR markers were eligible for NetBio this study. U.S. ANDE RapidHIT 200 Data transferred to NIST for analysis One chemistry and one instrument PowerPlex 16 Federal per set of 20 swabs Chemistry 5 PowerPlex 16 100 State NIST provided 20 single-NIST reporting success rate for all RapidHIT 200 One chemistry and one Data transferred to NIST 、 data/systems combined **GlobalFiler Express** source reference buccal IntegenX



Success was measured by complete and concordant genotypes produced by the integrated rapid DNA devices as compared to lab generated correct genotypes





Success rates indicate the average success for each STR locus group genotyped (CODIS 13 loci, PowerPlex 16 loci, new CODIS 20 loci). The minimum and maximum success rates observed within individual participants within the Maturity Assessment is represented by the whiskers of the boxplot above.



Private

		6

SlobalFiler	
Express	

60

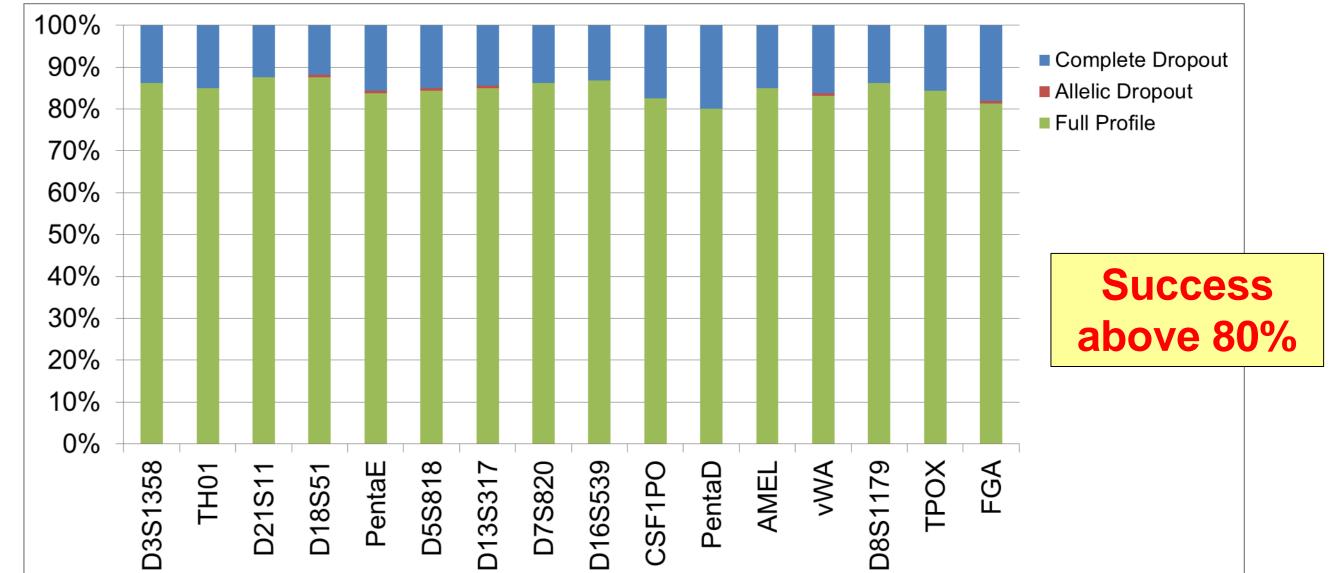
120

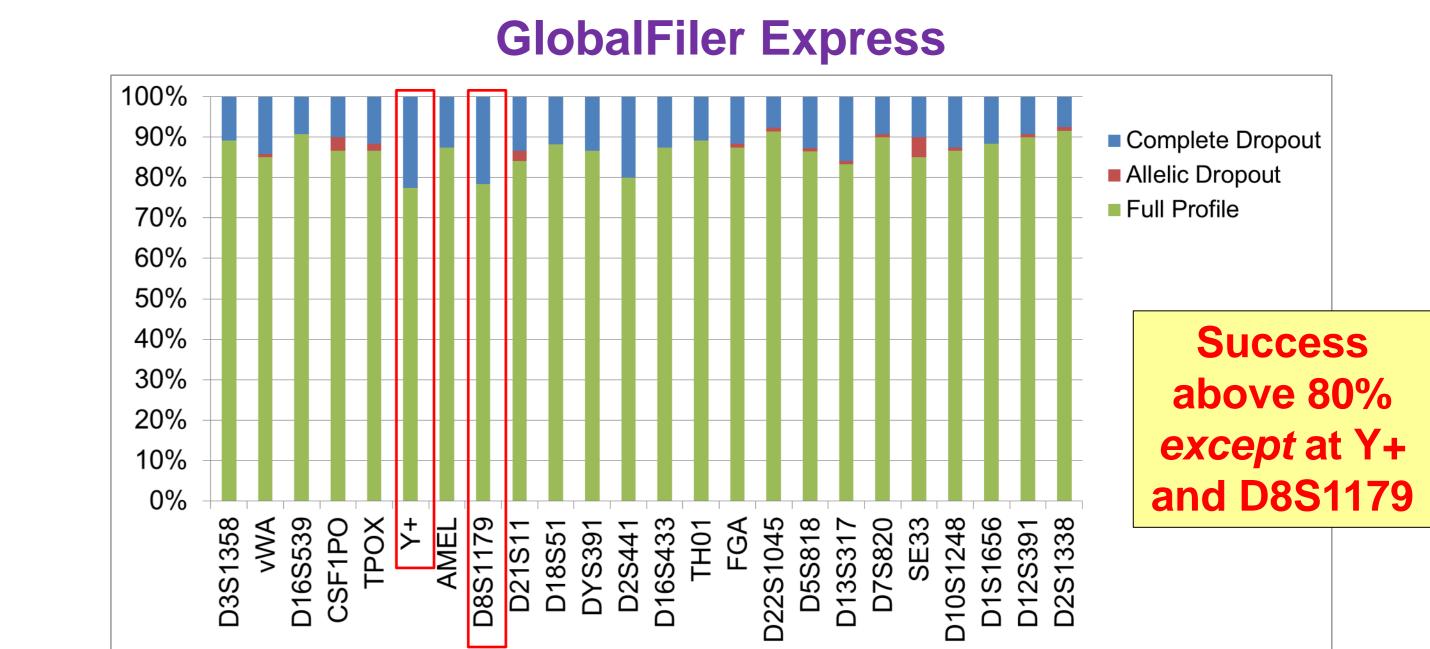
PowerPlex 16

Success: Per Locus

Percentage of successful genotypes generated per locus. The PowerPlex 16 data is a combination of the data generated from both ANDE and the RapidHIT 200.







Peak Height Ratios

Peak height ratios were calculated for all complete profiles the PowerPlex 16 and GlobalFiler Express chemistries.

Powerl	GlobalFiler Express					
Locus	Median			Locus	Median	
Penta E	0.81	40		SE33	0.79	
AMEL	0.83	50		D2S1338	0.82	
Penta D	0.84	40	_	D5S818	0.85	
D18S51	0.86		-	D18S51	0.85	
D3S1358	0.87		-	D12S391	0.86	
D8S1179	0.87		-	D21S11	0.87	
TPOX	0.87	м	-	CSF1PO	0.87	
D5S818	0.88	-	-	√WA	0.88	
WA VWA	-	-	D7S820	0.88		
VWA 0.88 D21S11 0.88				TPOX	0.89	
D16S539	0.88	-	-	D16S539	0.89	
D13S317	0.89	-	-	D1S1656	0.89	
CSF1PO	0.89		_	D22S1045	0.89	
FGA	0.89			D8S1179	0.90	
D7S820	0.89			D13S317	0.90	
TH01	0.93	-		AMEL	0.90	
				D3S1358	0.90	
n=118				D19S433	0.90	
The PowerPle	ata is a		D10S1248	0.91		
combination of the data				TH01	0.91	
				FGA	0.92	
generated fror		D2S441	0.92			
and the Rapid	0.	-		n=67		

		<u>S</u>	tutte	er Pe	rcen	tages	<u>S</u>	
for	Stutter p	ercentag	es wer	e calcula	ated for	all compl	ete pro	ofiles
	for the Po	owerPlex	k 16 an	id Globa	IFiler Ex	press ch	emistri	es.
PowerPlex 16 GlobalFiler Express						SS		
		Locus	Median		-	Loci	Median	
	-	Penta D	1.47		-	TH01	1.27	
	-		2.28	-	-	TPOX		-
	-	TPOX		4	-	D7S820		_
	_	Penta E		4	-	D2S441	4.76	_
	_	D7S820		4	-	DYS391	5.73	_
	_	D13S317		4	-	D16S539		-
	_	D18S51		4		D13S317		-
	-	D8S1179		+		CSF1PO		-
	_	CSF1PO		1	-	D8S1179		-
	_	D16S539		4	-	D18S51	6.67	-
	-		8.36	4	-	D5S818	6.76	
	_	D5S818		1	-	D22S1045	7.00	-
	-	FGA		4	-	D19S433		
	-	WWA		4	-	FGA		
	_	D3S1358		1	-	D3S1358		
		D21S11		4	-	D10S1248		_
n=118					-	D21S11	8.60	_
The PowerPlex 16 data is a					D2S1338			
combination of the data					D1S1656			
					٧WA	9.28	-	
generated from both ANDE					D12S391	9.46	-	
and the RapidHIT 200.				SE33	15.56			
							n=67	
							4.5	

All peak height ratios above 79%

Stutter within observed developmental validation range

Additional Resources

- https://www.fbi.gov/about-us/lab/biometric-analysis/codis/rapid-dna-analysis
- 2. http://www.swgdam.org/
- 3. https://www.fbi.gov/about-us/lab/biometric-analysis/codis/summary-of-rapid-dna-addenda-effective-12-1-14
- https://www.fbi.gov/about-us/lab/biometric-analysis/codis/rapid-dna-addendum-to-gas-final-effective-12-1-2014
- 5. https://www.fbi.gov/about-us/lab/biometric-analysis/codis/audit-document-for-rapid-dna-gas-addendum-effective-12-1-2014

Conclusions

Two fully integrated platforms were included in the 2014 Rapid DNA Maturity Assessment. A total of 11 instruments were tested between 7 laboratories for a total of 280 samples examined. Data for the Maturity Assessment was generated from October-December 2014 and returned to NIST for data analysis. Updates to instrumentation and software may have taken place since this Maturity Assessment was completed. Observed success for the CODIS 13 Core Loci ranged from 76% for Rapid DNA Analysis to 80% with Modified Rapid DNA Analysis.

Results can be located: http://www.nist.gov/mml/bmd/genetics/dna_biometrics.cfm

References:

1. Tan E, Turingan RS, Hogan C, Vasantgadkar S, Palombo L, Schumm JW, et al. Fully integrated, fully automated generation of short tandem repeat profiles. Investig. Genet. 4 (2013) 2041-2223.

2. Jovanovich, S., Bogdan, G., Belcinski, R., Buscaino, J., Burgi, D., Butts, E. L. R., et al. Developmental validation of a fully integrated sample-to-profile rapid human identification system for processing single-source reference buccal samples. Forensic Sci. Int. Genet. 16 (2015) 181-194.

3. Hennessy LK, Mehendale N, Chear K, Jovanovich S, Williams S, Park C, et al. Developmental validation of the GlobalFiler express kit, a 24-marker STR assay, on the RapidHIT System. Forensic Sci. Int. Genet. 13 (2014) 247-258.

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> Poster available for download from STRBase: http://www.cstl.nist.gov/biotech/strbase/pub_pres/RomsosISFG2015RapidDNA.pdf