# A Strategy for Characterization of Single Nucleotide Polymorphisms in a Reference Material



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The advent and adoption of next generation sequencing (NGS) is enabling analysis of single nucleotide polymorphisms (SNPs) at an unprecedented scale, limited primarily by multiplexing during the PCR amplification based enrichment step used for forensic applications. Since only a single nucleotide is assayed, PCR primers may be designed to generate small amplicons, making SNP markers well-suited to forensic DNA typing. Carefully selected panels of SNP markers have been previously established for forensic applications such as one-to-one matching, estimating biogeographical ancestry, and predicting externally observable phenotype [1,2,3,4,5,6]. To support the implementation of SNPs in forensic DNA analysis, NIST will examine the HID-Ion AmpliSeq Identity Panel and the HID-Ion AmpliSeq Ancestry Panel for the Ion Torrent Personal Genome Machine (PGM) and the Illumina ForenSeq DNA Signature kit for the Illumina MiSeq FGx. In total, over 300 SNP markers will be typed with approximately 40 % of loci being represented in more than one multiplex. A strategy combining NGS on orthogonal platforms and Sanger sequencing for characterizing the SNP markers for varying levels of confidence is presented herein. The outcome will be to report only the SNP allele calls, analogous to the mitochondrial sequence variants in NIST Standard Reference Material (SRM) 2392, and not the subsequent application of ancestry or phenotype). Additionally, we demonstrate the optimization of automated sequencing template preparation using the Ion Chef from Ion Torrent to create an efficient workflow suitable to the forensic DNA laboratory.

Specific Aim: To devise an efficient methodology for characterization of SNP genotype values in a reference material. Using NGS as a primary measurement method, concordance between NGS platforms may be used to obtain high confidence SNP genotype values without exhaustive Sanger sequencing. SNP genotypes discordant across platforms may be disambiguated with Sanger sequencing. Evaluation of Mendelian inheritance patterns in family trios may be used as an additional confirmatory strategy.

Next Generation Sequencing Platforms:	NIST Levels of Confidence for Certi	fication	Table 1: Genotypes of nine candidate reference materials characterized for           HID-Ion AmpliSeq Identity Panel (a) and the HID-Ion AmpliSeq Ancestry Pa					
Two platforms with commercial SNP genotyping panels intended for forensic DNA analysis are to be used in this	Description	Proposed Method	(b) using the Ion Torrent PGM sequence (a) PGM Identity Panel Locus RM 1 RM 2 RM 3 RM 4 RM 5 RM 6 RM 7 RM 8 RM 9	(b) PGM Ancestry Panel           Locus         RM 1         RM 2         RM 3         RM 4         RM 5         RM 6         RM 7         RM 8         RM 9				
characterization schema: the Ion Torrent Personal Genome Machine and the Illumina MiSeq FGx.	NIST has the highest confidence in its accuracy. All known or suspected	Sanger Sequencing + NGS	rs1490413       AA       AG       GG       AG       GG       AG       GG       AG       AA       AA       AA         rs7520386       AG       AG       GG       AG       GG       GG       AA       AA       AA       AA         rs4847034       GG       GG       AG       AG       AG       GG       AA       AA       AA       AA         rs560681       AG       AA       GG       AA       GG       AA       AG       AG         rs10495407       GG       AA       AG       GG       AA       AG       AG       AG         rs891700       AG       AA       AA       AG       AG       AG       GG       AG       AG       GG         rs1413212       CC       CC       CC       CC       CC       CC       CT       TT       TT         rs876724       CT       CT       CC       CT       CC       CT       CT       CT       TT	rs2986742         CC         TT         TT				

500ni	Continod	sources of bias have been investigated or taken into account.		rs1109037       AG       GG       GG       AA       GG       GG       AG       AA       rs7554936         rs93934       GG       AG       AA       AA       AG       AA       AG       AA       AG       AG       AA         rs12997453       AG       AG       GG       GG       GG       GG       AG       AA       AG       AG       AG       rs1040404       rs1040404       rs1040404       rs1040404       rs1407434       rs1407434       rs1407434       rs1407434       rs1407434       rs4951629       rs4364205       GT       GT       GG       GG       TT       GG       GG       GT       rs316873       rs316873       rs316873       rs316873
A A A A A A A A A A A A A A A A A	Reference	A high-confidence estimate of the true value but where all possible sources of bias have not been fully investigated by NIST.	NGS + NGS Two Platform Concordance	rs1872575         AG         GG         AG         AA         AG         AA         AA         AA         AA           rs1355366         TT         TT         TT         TT         CT         CT         TT         CC         CT         CC
Ion Torrent PGM         Credit: www.lifetechnologies.com    Illumina MiSeq FGx Credit: www.illumina.com	Informational	Data that may be of interest and use to the SRM user, but insufficient information is available to access the confidence of the assignment.	NGS on one platform	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$
Ion Torrent PGM				rs10776839         GG         GG         GG         GG         GT         TT         GG         GT         rs1007810         rs10007810         rs10007810         rs10007810         rs1369093         rs1369093         rs1369093         rs1369194         rs1385194         rs1385194 <thrs1385194< th=""> <thrs1385194< th=""> <t< td=""></t<></thrs1385194<></thrs1385194<>
HID-Ion AmpliSeq Identity Panel 90 Identity Informative SNPs (IISNPs) [1,2] 34 Lineage Informative SNPs (LISNPs) [3]	(a) 2 Standard Deviations 4000 MEAN	Sequencing Coverage for 124 IISNPs	Y-Chromosome SNPs	rs3780962       AG       IT
HID-Ion AmpliSeq Ancestry Panel 165 Ancestry Informative SNPs (AISNPs) [4,5]				rs1058083       GG       AG       GG       GG       GG       GG       AG       GG       AG       rs7722456         rs354439       AT       AT       T       TA       AA       AT       AT       TT       TT       rs6422347         rs1454361       AT       TT       AA       AT       AT       TT       TT       rs6422347         rs722290       CG       CC       GG       CG       CC       CG       CG       CG       rs1040045       rs2504853         rs873196       CT       TT       CT       CT       CT       CT       CT       TT       rs7745461       rs7745461         rs4530059       GG       AG       AA       GG       GG       GG       AG       AG       rs192655       rs192655       rs192655       rs192655       rs3823159       rs3823159       rs4453276       rs4453276       rs4453276       rs4453276       rs4453276       rs4453655
<u>Illumina MiSeq FGx</u>		Sequencing Coverage for 165 AISNIPs		rs729172         GT         TT         GG         GG         GT         GG         GG         GT         GG         GG         GT         rs1871428         rs1871428         rs1871428         rs731257         rs731257         rs731257         rs731257         rs731257         rs731257         rs1382387         CC         AA         AA         AA         AA         AC         AA         AG         rs917115         rs23214         rs23214         rs23214         rs232314         rs23214         rs232314         rs232314 <thr>         r         r         r</thr>
ForenSeq Signature Assay Kit - 172 SNP markers: 94 IISNPs [1,2] 56 AISNPs [5] 22 Phenotype Informative SNPs (PISNPs) [6]	(b) 2 Standard Deviations 3000	rs16891982 rs2306040	rs12439433 rs1296819	rs9905977       AG       GG       AG       GG       AG       GG       AA
Ion Torrent Illumina PGM MiSeg FGx				rs914165         AA         GG         AG         GG         GG         AG         GG         AG         AG         AG         AG         rs10511828         rs10511828         rs10511828         rs10511828         rs3793451         rs3793451         rs3793451         rs3793451         rs2306040         rs10511828
(a) Identity Informative SNPs	<b>Figure 2</b> : Sequencing coverage of nine with the 34 Y-chromosome LISNPs at th below two standard deviations of the me	candidate reference materials run in triplicate plotted for each e far right representing five male samples. (b) Ancestry Pane an coverage for all loci are circled.	ch SNP locus. (a) Identity Panel SNPs are shown el SNPs are shown. Loci with mean coverage	rs1020520       GG       AA       AG
HID-Ion ForenSeq	Indi	cators of Low Confidence Gend	otypes	P256       -       G       G       -       G       -       G       -       rs1730667         P202       -       T       T       -       T       -       T       -       rs1730667         rs17306671       -       A       T       -       A       -       A       -       rs1079597         rs17411895       -       A       -       A       -       A       -
Ampliseq408411SignatureIdentity PanelAssay	Sequencing coverage: The Ion Amplis protocol suggests a minimum sequencing	Seq Library Preparation for Human Identification App og coverage of 300 X for autosomal SNPs (150 X for Y roverage depth of 738 X is recommended for the Iden	olications (Pub. No. MAN0010640 Rev. A.0) Y-SNPs). To achieve this minimum coverage	ISH41000       -       A       A       -       A       A       -       ISH48028       - </td



Figure 1: Venn diagrams of the SNP loci in Ion Torrent and Illumina NGS multiplexes with (a) overlapping identity informative markers (IISNPs) and (b) ancestry/phenotype informative markers (AISNPs). The Ion Torrent panels do not include any Phenotype Informative SNPs. There are 139 loci in common (out of a total 323) between the two platforms.

### Methods:

Nine single-source DNA samples have been selected as candidate components for a reference material.

Name	Description
RM 1	Female
RM 2	Male
RM 3	Male
RM 4	Female
RM 5	Male
RM 6	Female
RM 7	Family Trio, Son
RM 8	Family Trio, Father
RM 9	Family Trio, Mother

- Typed on PGM Identity Panel & Ancestry Panel
- DNA input  $\approx$  1 ng
- Three replicate amplifications per sample
- Used library protocol for PGM HID SNPs
- ISP templating & chip loading performed on lon Chef

These recommended values allow for locus-to-locus variations in sequencing coverage balance. This translates to a maximum of 77 samples in a library pool when using an Ion 318 Chip for the Identity Panel or 59 samples when running the Ancestry Panel, assuming 80 % chip loading and 60 % usable reads. During NIST's pilot PGM sequencing experiments discussed herein, 31 samples were sequenced concurrently on one 318 chip for each panel. This is roughly half of the maximum number of samples recommended for a 318 chip. In this dataset, two (2) out of 2430 (0.08 %) Identity Panel autosomal SNP genotypes had sequencing coverage below 300 X. The number of Identity Panel Y-chromosome LISNPs with coverage below 150 X was one (1) out of 597 (0.17 %). In the Ancestry Panel data, 90 SNP genotypes out of a total 4455 (2.02 %) were below the suggested minimum 300 X coverage. Developmental validation criteria discussed in the protocol require that the average coverage for a locus should be within two standard deviations of the mean coverage for all loci. Here the mean value of Identity Panel autosomal IISNP loci is 1747.3 X +/- 531.5 X, making the lower threshold for coverage 684.3 X. For Ychromosome LISNPs, the mean was 849.5 X +/- 236.4 X, making the lower cutoff 376.6 X. For the Ancestry Panel the mean value was 1063.1 X +/- 354.0 X, making the lower threshold for coverage 355.1 X. Higher sequencing coverage was not considered a risk factor for

low confidence measurements. Following this principle, one autosomal locus in the Identity Panel (rs2342747 @ 466.1 X) and one Y- Table 2: Average strand bias chromosome locus (M479 @ 280.2 X) fell below two standard deviations of the mean, while five Ancestry Panel loci were below the criterion: rs1296819 (284.9 X), rs2306040 (323.9 X), rs12439433 (326.0 X), rs16891982 (332.4 X), rs13400937 (346.4 X). Loci with characteristically low coverage are highlighted with orange circles in Figure 2 and with orange filled cells in Table 1.



(% positive strand) for loci with at least one data point < 35 % or > 65 %



Figure 3: Allelic balance of nine single-source samples run in triplicate plotted for each SNP locus. (a) Identity Panel SNPs are shown with the 34 Ychromosome LISNPs at the far right, representing five male samples. All Y-chromosome LISNPs are homozygous. (b) Ancestry Panel SNPs are shown. Samples with average allelic balance > 65 % or < 95 % are circled in blue.

Allelic imbalance: heterozygotes should have a sequencing read ratio of 50 % and homozygotes should be 100 %. Deviation from these values may be an indication that a technical issue has occurred with the determination of the genotype. Heterozygotes deviating from the expected 50 % (+/- 15 %) allele ratio may have a SNP underlying a PCR primer binding site which disturbs PCR efficiency or other technical issue such as strand bias or systematic sequencing error. Homozygotes exhibiting a ratio below 100 % (+/- 5 %) could be an indication of systematic sequencing error or bioinformatic sequence alignment issue. Loci with imbalanced allelic ratios are summarized in Table 3. A previous study be by Seo et al. [7] noted that marker rs4530059 in an earlier version of the Identity Panel displayed allelic imbalance in several heterozygous individuals where the average coverage ratio of A/G was 67.6 %.



	rs4411548					11				
	rs2593595	AA	AA	AA	AG	AA	AA	AA	AA	AG
	rs17642714	AT	AA	AA	AA	AA	AA	AT	AA	AT
	rs4471745	GG	GG	AA	GG	AG	GG	GG	GG	GG
	rs2033111	AA	AG	AA	AG	AG	AA	AA	AA	AA
	rs11652805	Π	СТ	СТ	Π	Π	TT	Π	Π	Π
	rs10512572	GG	AG	AG	GG	GG	GG	GG	GG	GG
I	rs2125345	CC	Π	TT	СС	СТ	TT	Π	Π	СТ
I	rs4798812	GG	AG	GG	AG	AG	GG	AA	AG	AA
I	rs2042762	Π	СТ	TT	TT	Π	TT	TT	Π	TT
I	rs7226659	GG	GT	TT	GG	GG	GG	GG	GG	GG
I	rs7238445	AG	GG	GG	GG	AG	GG	GG	GG	GG
I	rs881728	CC	CC	AC	AC	CC	CC	AC	AC	CC
I	rs3916235	СС	CC	CC	СС	CC	CC	СТ	СТ	CC
I	rs4891825	AA	AA	AA	AA	AG	AA	AG	AG	AA
I	rs874299	СТ	СТ	TT	СТ	СТ	СТ	CC	CC	CC
	rs7251928	AA	AC	AC	AA	AA	CC	AC	AC	AA
	rs8113143	AC	AC	AC	AC	AC	AC	CC	CC	CC
	rs3745099	AA	AG	AG	AA	AA	AG	AA	AG	AG
	rs2532060	СТ	CC	CC	СС	CC	TT	СТ	СТ	СТ
	rs6104567	Π	GG	TT	GT	GT	GT	TT	GT	TT
	rs3907047	Π	СТ	TT	Π	Π	TT	СТ	Π	СТ
	rs310644	Π	Π	CC	Π	Π	TT	СТ	СТ	TT
	rs2835370	Π	Π	TT	TT	Π	TT	СТ	СТ	TT
	rs1296819	CC	CC	AC	CC	AC	CC	AA	AC	AC
	rs4821004	СТ	СТ	СТ	CC	СТ	CC	CC	CC	CC
	rs2024566	AG	AA	AA	AA	AA	AG	AA	AA	AA
	rs5768007	CC	СТ	CC	СС	CC	CC	CC	CC	СС

Table 3: Genotypes and allelic ratios of loci exhibiting allelic imbalance (shaded blue) for the HID-Ion AmpliSeq Identity Panel (a) and HID-Ion AmpliSeq Ancestry Panel (b). \* Strand bias observed for this locus.

I	Locus	RM 1	RM 2	RM 3	RM 4	RM 5	RM 6	RM 7	RM 8	RM 9
	rs7520386	AG	AG	GG	AG	GG	AA	AA	AA	AA
	Ratio (%)	71.6 +/- 1.5	70.3 +/- 0.7	99.9 +/- 0.1	71.2 +/- 5.2	99.8 +/- 0.3	99.8 +/- 0.1	99.9 +/- 0.0	99.8 +/- 0.0	99.9 +/- 0.0
I	rs4530059	GG	AG	AA	GG	GG	GG	AG	AG	GG
	Ratio (%)	99.9 +/- 0.1	70.4 +/- 2.9	99.5 +/- 0.1	99.8 +/- 0.2	99.9 +/- 0.1	100.0 +/- 0.0	73.4 +/- 0.5	77.9 +/- 7.4	99.9 +/- 0.1
	rs430046 *	СТ	СТ	CC	Π	СТ	СТ	СТ	СТ	СТ
I	Ratio (%)	66.7 +/- 3.6	64.2 +/- 0.4	100.0 +/- 0.0	99.9 +/- 0.0	64.4 +/- 2.3	63.7 +/- 1.6	65.9 +/- 1.7	63.9 +/- 5.5	65.8 +/- 3.7
	(1, )									

Locus	RM 1	RM 2	RM 3	RM 4	RM 5	RM 6	RM 7	RM 8	RM 9
rs734873	GG	GG	GG	GG	GG	GG	AG	AG	AG
Ratio (%)	100.0 +/- 0.0	100.0 +/- 0.1	100.0 +/- 0.0	100.0 +/- 0.1	100.0 +/- 0.0	99.9 +/- 0.0	55.4 +/- 4.7	68.2 +/- 4.5	53.2 +/- 2.7
rs7722456	TT	Π	СС	Π	TT	Π	TT	Π	TT
Ratio (%)	93.5 +/- 1.6	95.2 +/- 1.0	100.0 +/- 0.1	92.8 +/- 0.9	91.6 +/- 0.5	93.3 +/- 1.7	91.7 +/- 2.2	91.3 +/- 3.0	91.3 +/- 1.4
rs3943253	AA	AG	AA	AA	AG	AG	AA	AA	AG
Ratio (%)	99.9 +/- 0.1	54.3 +/- 1.6	93.2 +/- 1.4	99.9 +/- 0.1	51.0 +/- 1.3	51.2 +/- 1.0	99.8 +/- 0.1	99.9 +/- 0.1	51.6 +/- 0.6
rs4918664	AA	GG	GG	AA	AA	AG	AA	AA	AA
Ratio (%)	99.7 +/- 0.4	94.8 +/- 0.5	99.8 +/- 0.2	99.8 +/- 0.2	100.0 +/- 0.1	52.8 +/- 2.6	99.9 +/- 0.1	99.9 +/- 0.0	100.0 +/- 0.0
rs2899826	AA	GG	GG	AA	AA	AA	AA	AA	AA
Ratio (%)	99.8 +/- 0.1	91.3 +/- 0.7	90.4 +/- 0.7	99.8 +/- 0.2	99.8 +/- 0.1	99.9 +/- 0.1	99.9 +/- 0.1	99.9 +/- 0.1	99.9 +/- 0.1
rs7251928 *	AA	AC	AC	AA	AA	CC	AC	AC	AA
Ratio (%)	99.7 +/- 0.2	86.2 +/- 2.0	67.7 +/- 3.3	99.9 +/- 0.1	99.8 +/- 0.1	88.4 +/- 1.8	67.5 +/- 1.0	67.5 +/- 2.5	99.8 +/- 0.1

#### **Discussion:**

- 31 samples in barcoded library pool
  - 9 candidate RMs + 1 control in triplicate
- One negative control
- Sequenced each panel on one Ion 318 Chip

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Strand bias: defined as the number of plus strand reads divided by the number of minus strand reads, this condition indicates a systematic sequencing error type wherein one strand yields low quality results and is bioinformatically filtered from the final data. Using an arbitrary cutoff of +/- 15 % from the expected ideal ration of 50 % to demark low confidence genotypes, there were five loci in the Identity Panel and nine loci in the Ancestry Panel which yielded at least one data point outside the arbitrary limits of < 35 % and > 65 %. These SNP markers are summarized in Table 2. Data points falling outside these arbitrary thresholds may be considered low confidence and will require confirmation with a secondary method.

Discordant replicates: two loci in the Ancestry Panel had discordant genotypes between triplicate data points (see yellow shaded cells in Tables 1 & 3). For marker rs3943253 one replicate had a no-call (NN) while the two remaining replicates were homozygous (AA). This marker has an allelic ratio of 93.2 % for the two successfully genotyped replicates and 90.3 for the no-call, however, no quality flag was issued to explain the no-call. All replicates of rs3943253 had some proportion of reads supporting a "G" genotype, which is the alternate allele; the adjacent bases are 5' T and 3' C. The G impurity appears in both forward and reverse reads. No clear explanation is available for this mis-genotype. It may be a case of context-specific sequencing error. A second locus, rs7251928, had one replicate with genotype "CC" and two with "AC". Preliminary data from an alternate platform suggests that the correct call for this locus is "CC". Given that all heterozygotes displayed notable strand bias and allelic imbalance at this locus, resulting genotypes should be considered "very low confidence" until confirmed with an alternate method.

**Mendelian inheritance patterns:** family trio samples may be used to verify proper genotyping performance. Maternal and Paternal allele inheritance patterns are depicted in **Table 4 below**. Nonconformities in expected inheritance patterns may indicate a technical issue with genotyping a locus. This approach may also be used to identify loci which contain putative SNPs underlying primer binding regions. No inconsistencies in inheritance patterns were identified among family trio samples in the candidate reference materials when evaluated with the Identity Panel or Ancestry Panel. Due to the criteria used in selecting the loci for the Identity Panel (high heterozygosity, low F<sub>st</sub>) and the Ancestry Panel (low heterozygosity, high F<sub>st</sub>), there are differences in the percentages of autosomal alleles which may be definitively determined to have originated from one parent versus the other. In the Identity Panel, 38.9 % of loci were informative versus 20.6 % for the Ancestry Panel.

The majority of SNP loci in the Ion Torrent Identity and Ancestry Panels satisfy proposed requirements for an "informational" classification in a NIST SRM. A small number of loci in these Panels were found to have characteristics signifying potential technical sources of error leading to uncertainty in the measurement. These markers require additional characterization before achieving any status. Confirmation of SNP genotypes with a second NGS method would satisfy the proposed requirements for a "reference value" in a NIST SRM. Final confirmation with Sanger-type sequencing is required to achieve a "certified" value under the proposed framework.

## **Future Directions:**

There are 139 loci which are present in both the HID-Ion AmpliSeq Panels and the Illumina ForenSeq Signature Prep kit. Concordance between the two platforms would qualify those loci as "reference values" for a NIST SRM. Discordant loci will require disambiguation using an alternate method such as Sanger sequencing in order to achieve any status for a NIST SRM. NIST intends to characterize the SNP loci in the Illumina ForenSeq Signature kit. NIST will consider the needs of the forensic community in terms of required levels of confidence in genotypes when prioritizing tasks leading to the potential development of a reference material for SNP genotyping.

Table 4: Mendelian inheritance patterns in Family trio samples for the HID-Ion AmpliSeq Identity Panel SNPs (a) and HID-Ion AmpliSeq Ancestry Panel SNPs (b)

(a)

80

70

(b)

