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STRBase, Textbooks, and NIST Research Efforts: Developing Tools to Aid the Forensic DNA Community

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Presentation Topics

- Introduction to NIST and to Our Group
- STRBase website
- Textbooks
 - Advanced Topics in Forensic DNA Typing (3rd edition)
- Group Research Overview
 - Standard Reference Materials (SRMs)
 - STR loci characterization
 - STR kit concordance studies BECKY
 - ABI 3500 validation work ERICA
 - TrueAllele software examination MIKE
 - Rapid PCR amplification PETE
- ABI 3500 open letter status update

NIST Human Identity Project Teams within the Applied Genetics Group

Guest

Researcher

Forensic DNA Team

Funding from the National Institute of Justice (NIJ) through NIST Office of Law Enforcement Standards





John Butler

Mike Coble

Becky Hill

Workshops & Textbooks

Concordance & LT-DNA Mixtures. mtDNA & Y

SRM work. variant alleles & Cell Line ID

Margaret

Kline

Data Analysis

Alvarez



Dave Duewer

DNA Biometrics Team

Funding from the FBI S&T Branch through NIST Information Access Division



Pete Vallone



Butts

Kristen Lewis O'Connor

Rapid PCR. Direct PCR & Biometrics

ABI 3500 & DNA Extraction D12/vWA & Kinship Analysis



http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm



National Institute of Standards & Technology (NIST)

- Non-regulatory agency established in 1901 in the US Department of Commerce.
- Mission to promote US innovation and industrial competitiveness by advancing measurement science, standards & technology.
- NIST develops a wide variety of physical standards, test methods, and standard reference data.



NIST Gaithersburg Campus



NIST Biochemical Science Division





NIST Applied Genetics Group

Group Leader











John **Butler**

Marcia Holden

Margaret **Kline**

Pete Vallone

Mike Coble





Hill





Erica **Butts**





Kevin Kiesler





APPLIED GENETICS Group

Major Programs Currently Underway

Forensic DNA

- STRBase website
- New loci and assays (26plex)
- STR kit concordance
- Ancestry SNP assays
- Low-template DNA studies
- Mixture interpretation research and training
- STR nomenclature
- Variant allele cataloging and sequencing
- Expert systems review
- Training workshops to forensic DNA laboratories
- Validation experiments, information and software tools
- **Textbooks** 3rd ed. (3 volumes)

- Clinical Genetics
 - Huntington's Disease SRM
 - CMV SRM
 - Exploring future needs

Ag Biotech

 – "universal" GMO detection/ quantitation (35S promoter)

DNA Biometrics

- Rapid PCR methods
- Efforts to standardize testing of future portable DNA systems
- Kinship analysis
- **Cell Line Authentication**

STRBase

NIST STRBase Website: Forensic DNA Information Resources

http://www.cstl.nist.gov/biotech/strbase/



Internet DataBase

NIST Standard Reference Database SRD 130 [Recent Updates]

Serving the forensic DNA and human identity testing communities for over 10 years... These data are intended to benefit research and application of short tandem repeat DNA markers to human identity testing. The authors are solely responsible for the information herein.

This database has been accessed 368383 times since 10/02/97. (Counter courtesy www.digits.com - see disclaimer.)

Created by <u>John M. Butler</u> and <u>Dennis J. Reeder</u> (<u>NIST Biochemical Science Division</u>), with invaluable help from Jan Redman, Christian Ruitberg and Michael Tung Site creators' curriculum vitaes available using links above.

I invite your lab to supply information on variant and tri-alleles observed

History of STRBase Website

- Information was collected on STR markers while working on my PhD dissertation in 1993-1995
- Started a review article in 1996 while a NIST postdoc but wanted to create a dynamic rather than an out-of-date resource
- Created hundreds of individual web pages that were hyperlinked together
- Website launched in July 1997
- I continue to update the website (via an HTML editor)...

New STRBase Sections

Forensic STR Information

- STRs101: Brief Introduction to STRs
- o Core Loci: FBI CODIS Core STR Loci and European Core Loci
- STR Fact Sheets (observed alleles and PCR product sizes)
- Multiplex STR kits
- o Sequence Information (annotated)
- Variant Allele Reports
- <u>Tri-Allelic Patterns</u>
- Mutation Rates for Common Loci
- <u>Published PCR primers</u>
- o <u>Y-chromosome STRs</u>
- Low-template DNA Information Updated
- o Mixture Interpretation
- o Kinship Analysis
- miniSTRs (short amplicons) ◆
- <u>Null Alleles</u> discordance observed between STR kits
- STR Reference List now 3400 references

NIST Standard Reference Family Pedigree



Data available for testing software programs: http://www.cstl.nist.gov/biotech/strbase/kinship.htm

Textbooks

Forensic DNA Typing Textbook

1st Edition



Jan 2001 335 pp. 17 chapters



With Y. Fukuma (Japanese translator)

2nd Edition



Feb 2005 688 pp. 24 chapters

Now available in **Chinese** (Yiping Hou)

Japanese in preparation (Yoshiya Fukuma)



Sept 2009



Sept 2011



3rd Edition

With Y. Hou (Chinese translator)

Forensic DNA Typing Textbook 3rd Edition is Three Volumes



Sept 2009



Advanced Topics in Forensic DNA Typing: INTERPRETATION

Sept 2011

Fall 2012

~500 pages

~700 pages

~500 pages

New Materials in Advanced Topics book Released August 2011

- Cites >1500 new references (>2800 ref. total)
- New chapter on legal aspects

 expert witness prep, perspectives from lawyers
- New chapter on X-chromosome markers
- Extensive updates on LCN, Y-STRs, miniSTRs, mtDNA, SNPs, non-human DNA, and database issues
- Coverage of all the new STR kits
- Listing of all known STR alleles for all 23 kit loci

J.M. Butler (2011), Advanced Topics in Forensic DNA Typing: Methodology, Figure 5.5		2	= 13 CODIS STR Loci							Additional 10 STRs						;									
ے Autosomal STR Typing Kits		Amelogeni	CSF1P0	FGA	TH01	трох	vWA	D3S1358	D5S818	D7S820	D8S1179	D13S317	D16S539	D18S51	D21S11	D2S1338	D19S433	Penta D	Penta E	D1S1656	D2S441	D10S1248	D12S391	D22S1045	SE33
	ABI AmpFISTR kits																								
	Profiler	1	4	3	2	3	2	1	1	3		2													
	Profiler Plus (ID)	1		3			2	1	1	3	2	2		4	3										
	COfiler	1	4		2	3		1		1			2												
	SGM Plus	1		3	2		2	1			2		3	4	3	4	1								
10	Identifiler (Direct, Plus)	1	4	3	2	3	2	1	2	3	1	3	4	4	2	5	1								
N.	SEfiler Plus	1		3	2		2	1			2		3	2	1	4	1								3
	MiniFiler	1	1	2						2		1	1	2	3	2									
X	NGM	1		4	3		2	2			2		3	4	3	4	2			3	1	1	4	1	
	NGM SElect	1		4	3		2	2			2		3	4	3	4	2			3	1	1	4	1	5
	Promega PowerPlex kits		_							•		0											T		
ō	PowerPlex 1.1 (1.2)	3	5		2	4	1		1	3	0	2	4						-						
		4	_	4	2	3	1	1	4	0	2	0	4	4	3			0	5						
	PowerPlex 16 (BIU, HS)	1	5	5	2	4	2	1	1	3	3	2	4	4	3			6	5						
	PowerPlex S5	1		2	1		0	4			2			3											1
N		1		4	2		2	1			3 2		4	4	3	0	2			2	1	4	2	1	
	PowerPlex ESX 16	1		4	3		2	2			3		4	5	4	3	3			2	1	1	2		4
	PowerPlex ESA 17	1		4	3		2	2			3		4	5	4	3	3			2	 	1	2	 	4
S	PowerPlex ESI 10	1		2	1		2	2			1		1	2	3	4	<u>ა</u>			ა ი	ວ 	4	4	ວ 	2
	PowerPlex ESI 17	1	5	2	2	Δ	2	∠ 1	1	2	1 2	2	1	2	3	4	о 1	G	F	<u>ی</u>	Э	4	4	ວ	<u>ు</u>
3	Oiagan Investigator kits	1	J	J	2	4	2	l		3	<u></u>	2	4	4	3	2		0	3						
	Eggnlav	1		4	2		4	3			Δ		1	2	5	5	3			2	1	1	3	2	
	ESSNER SE	1		3	2		4	3			4		1	2	5	5	3			2	1	1	3	2	4
	Hexaplex FSS	1			1						-			2	5	5	0			3	1	2	2	2	
	Nonaplex ESS	1		4	2		4	3			2			1	5					2	1	1	3	2	3
	Decaplex SF	1		2	2		4	3			3		1	1	5	4	2			_					1
	IDplex	1	2	4	2	1	4	3	4	2	2	3	1	1	5	5	3								

Common Forensic STR Loci



Jnited States



The 10 STR Loci Beyond the CODIS 13

	STR Locus	Location	Repeat Motif	Allele Range*	# Alleles*
	D2S1338	2q35	TGCC/TTCC	10 to 31	40
	D19S433	19q12	AAGG/TAGG	5.2 to 20	36
	Penta D	21q22.3	AAAGA	1.1 to 19	50
	Penta E	15q26.2	AAAGA	5 to 32	53
loci	D1S1656	1q42	TAGA	8 to 20.3	25
ean	D12S391	12p13.2	AGAT/AGAC	13 to 27.2	52
urop	D2S441	2p14	TCTA/TCAA	8 to 17	22
N E	D10S1248	10q26.3	GGAA	7 to 19	13
5 ne	D22S1045	22q12.3	ATT	7 to 20	14
	SE33	6q14	AAAG [‡]	3 to 49	178

*Allele range and number of observed alleles from Appendix 1, J.M. Butler (2011) Advanced Topics in Forensic DNA Typing: Methodology; [‡]SE33 alleles have complex repeat structure

SE33 (58 alleles observed)

	Total Populations, %							Total Populations, %					
Allele	#	%	Af Am	Asian	Cauc	Hisp	Allele	#	%	Af Am	Asian	Cauc	Hisp
6.3							23	12	0.4	0.6	1.0	0.2	0.1
7	34	3 a	enoty	bes o	obse	rved	23.2	91	3.2	2.2	4.2	4.3	2.1
8		3	·····,				24	1	0.0			0.1	
10.2	H	eter	ozygo	DSITY	= 0.9	311	24.2	74	2.6	1.3	6.2	2.2	2.5
11		0.0			0.1		25.2	109	3.8	2.6	6.9	4.0	3.1
11.2	2	0.1	0.2				26	1	0.0	0.1			
12	11	0.4	0.3		0.5	0.4	26.2	163	5.6	6.1	5.2	4.3	7.1
12.2	4	0.1	0.2			0.3	27	1	0.0				0.1
13	31	1.1	1.1		1.5	1.0	27.2	225	7.8	4.3	10.4	9.5	8.6
13.2	9	0.3	1.0	0.0	0.5		27.3	2	0.1				0.3
14	85	2.9	5.1	0.2	2.5	2.4	28	2	0.1	0.1	0.2		
14.2	10	0.3	0.4	1 0	0.4	0.3	28.2	180	6.2	4.4	7.9	7.4	6.1
15 2	102 8	3.3 0.3	3.9 0.3	1.2	3.9	3.9	28.3	2	0.1	0.1		0.1	
16	144	0.3 5 0	0.5	47	4.0	67	29	1	0.0		0.2		
16.2	5	0.2	0.3	7.7	4.0 0 1	0.7	29.2	147	5.1	2.7	5.7	6.3	6.3
16.3	2	0.1	0.0		0.1	0.3	29.3	1	0.0		0.2		
17	205	7.1	9.3	4.0	6.2	7.3	30	1	0.0				0.1
17.2	1	0.0	0.1				30.2	111	3.8	1.6	3.2	5.8	4.6
17.3	5	0.2	0.1		0.2	0.3	31	3	0.1	0.1		0.2	
18	268	9.3	12.1	5.0	7.2	11.0	31.2	52	1.8	1.5	2.5	2.2	1.3
18.3	1	0.0			0.1		32	1	0.0			0.1	
19	250	8.7	12.4	6.2	6.6	8.0	32.2	25	0.9	0.4	0.7	1.3	0.9
19.2	8	0.3	40.0	0.2	0.4	0.4	33	2	0.1			0.1	0.1
20	216	1.5	10.9	9.2	5.4	4.8	33.2	11	0.4	0.3		0.5	0.4
20.2	20	0.7	0.3	1.2	1.1	0.3	34	9	0.3	0.3		0.7	
21	801 مر	3.1 1 7	4.0	0.1 1 7	Z.4	2.1	34.2	1	0.0			0.1	
21.2 22	40 12	1.7	1.1	1./	2.4 1 5	1.0	35	1	0.0	0.1		-	
22.2	+∠ 65	2.3	0.4	3.2	3.8	1.9	36	2	0.1	0.2			
				U . 	0.0								

Loci sorted on Probability of Identity (P _I) values									
	Alleles	Genotypes	Het.	P _I value					
STR Locus	Observed	Observed	(obs)	N = 938					
SE33	53	292	0.9360	0.0069					
Penta E*	20	114	0.8799	0.0177					
D2S1338	13	68	0.8785	0.0219					
D1S1656	15	92	0.8934	0.0220 -					
D18S51	21	91	0.8689	0.0256					
D12S391	23	110	0.8795	0.0257					
FGA	26	93	0.8742	0.0299					
Penta D*	16	71	0.8754	0.0356					
D21S11	25	81	0.8358	0.0410					
D19S433	16	76	0.8124	0.0561					
D8S1179	11	45	0.7878	0.0582					
vWA	11	38	0.8060	0.0622					
D7S820	11	32	0.8070	0.0734					
TH01	8	24	0.7580	0.0784					
D16S539	9	28	0.7825	0.0784					
D13S317	8	29	0.7655	0.0812					
D10S1248	12	39	0.7825	0.0837					
D2S441	14	41	0.7772	0.0855					
D3S1358	11	30	0.7569	0.0873					
D22S1045	11	42	0.7697	0.0933 -					
CSF1PO	9	30	0.7537	0.1071					
D5S818	9	34	0.7164	0.1192					
ΤΡΟΧ	9	28	0.6983	0.1283					

23 STR Loci present in STR kits rank ordered by their variability

Better for mixtures (more alleles seen)

There are several loci more polymorphic than the current CODIS 13 STRs

Better for kinship (low mutation rate)

NIST Projects

A Short Overview...

Congress Passed the DNA Identification Act of 1994 (Public Law 103 322)

Formalized the FBI's authority to establish a national DNA index for law enforcement purposes.

FBI's DNA Advisory Board *Quality Assurance Standards* **for Forensic DNA Testing Laboratories**



(October 1, 1998)

STANDARD 9.5

The laboratory shall check its DNA procedures annually or whenever substantial changes are made to the protocol(s) against an appropriate and available NIST standard reference material or standard traceable to a NIST standard.

NIST DNA Reference Materials

Date of release or certificate revision (r)

Forensic Applications

- STR PCR DNA Profiling (SRM 2391c) 1995, r2008, 2011
- Mitochondrial DNA Sequencing (SRM 2392-I, 2392) 1999, 2003, r2009
- Human Y-Chromosome DNA Profiling (SRM 2395) 2003, r2008
- RFLP DNA Profiling (SRM 2390) 1992, r2001, now obsolete

Clinical Applications

- Fragile X Human DNA Triplet Repeat (SRM 2399) 2004, r 2007
- Huntington's Disease CAG Repeats (SRM 2393) 2011
- Cytomegalovirus (SRM 2366) 2011

Platform Testing

- Human DNA Quantitation (SRM 2372) 2007
- Heteroplasmic mtDNA Mutation Detection (SRM 2394) 2004
- DNA Sequence Library for External RNA Controls (SRM 2374) in process

A few others are in early stages of development



Steps Involved in SRM Production

Attend conferences, read the literature, talk to potential customers

Sequence & Copy Number









Receive input on priorities for projects and potential SRMs

Research potential properties and samples to be characterized and measurement method to be used

Obtain candidate components/make measurements

Decide on number of SRM units to produce (impacts price/unit), sample packaging, concentration, etc.

Bottle components and conduct homogeneity and stability studies; finalize uncertainty

Write Report of Analysis and Certificate of Analysis

Certificate Reviewed and Approved by NIST Measurement Services Division

SRM Made Available for Purchase http://www.nist.gov/srm

Bottling SRM 2372 Materials



Teflon container holding ≈ 250 mL of *Candidate* SRM 2372. *It's not an SRM until it passes all testing.*

With a multi-channel pipettor 8 tubes can be filled at a time. That's \approx 214 reps to fill 1700 tubes per component.

The assembly line closing the recently filled tubes



Protecting the SRM Product from the Staff : Lab Coats, Masks and Hair nets or full face shields **P**ersonal **P**rotective Equipment (PPE) or **P**roduct **P**rotective Equipment.

Safety Considerations: The Blister Brigade



Closing the 1,700 component A tubes (SRM 2372) caused some blisters even while wearing gloves.

Safety resolution: Band-aids applied prior to closing SRM component tubes the next session helped reduce the number of blisters formed!

NIST Standard Reference Material (SRM) for Forensic DNA Testing

SRM 2391b (2003-2011)

- 48 autosomal STR loci with certified values
- **10 liquid genomic DNA** components + **2 punches** (cells on 903 paper)
- All single source samples
- 4 males + 6 females
- 9947A & 9948 included

- 23 autosomal STR loci and 17 Y-STRs certified
- 4 liquid genomic DNA components + 2 punches (cells on FTA & 903 paper)
- 5 single source + 1 mixture
- 3 males + 2 females (unique)
- All new samples
 - no 9947A or 9948

SRM 2391c to replace SRM 2391b and SRM 2395 (for Y-STRs)

SRM 2391c (2011-future)

Commercially Available STR Kits

Applied Biosystems (17)

- AmpFISTR Blue (1996)
- AmpFISTR Green I (1997)
- Profiler (1997)
- Profiler Plus (1997)
- COfiler (1998)
- SGM Plus (1999)
- Identifiler (2001)
- Profiler Plus ID (2001)
- <u>SEfiler (2002)</u>
- Yfiler (2004)
- MiniFiler (2007)
- SEfiler Plus (2007)
- Sinofiler (2008) China only
- Identifiler Direct (2009)
- NGM (2009)
- Identifiler Plus (2010)
- NGM SElect (2010)

Promega Corporation (13)

- PowerPlex 1.1 (1997)
- PowerPlex 1.2 (1998)
- PowerPlex 2.1 (1999)
- **PowerPlex 16** (2000)
- PowerPlex ES (2002)
- PowerPlex Y (2003)
- PowerPlex S5 (2007)
- **PowerPlex 16 HS** (2009)
- PowerPlex ESX 16 (2009)
- PowerPlex ESX 17 (2009) •
- PowerPlex ESI 16 (2009)
- PowerPlex ESI 17 (2009)
- PowerPlex 18D (2011)

Qiagen (2010)

Primarily selling kits in Europe Due to patent restrictions cannot sell in U.S.

- ESSplex
- ESSplex SE
- Decaplex SE
- IDplex
- Nonaplex ESS
- Hexaplex ESS
- HD (Chimera)
- Argus X-12
- Argus Y-12
- DIPlex (30 indels)

~1/3 of all STR kits were released in the last two years

STR Kits Tested with SRM 2391c

Applied Biosystems (12)

Identifiler Identifiler Plus NGM NGM SElect COfiler Profiler Profiler Plus Profiler Plus ID SGM Plus SEfiler Plus MiniFiler Yfiler Promega (9) PowerPlex 16 PowerPlex 16 HS PowerPlex Y PowerPlex ESX 17 PowerPlex ESI 17 PowerPlex S5 PowerPlex Y FFFL Qiagen (2) ESSplex IDplex <u>NIST assays</u> 26plex miniSTRs

Alleles sequen	ced:
SE33	
D12S391	
D1S1656	
Penta D	
Penta E	
D8S1115	

23 commercial STR kits examined NIST developed 26plex and miniplexes <u>No discordant results</u> observed on SRM 2391c samples

Kit Concordance Comparisons

Kits compared	<u>Samples</u>	Loci compared	Comparisons	# Differences	Concordance (%)
SGM-ID	1436	11	15,796	1	99.994
ID-ProPlus	1427	10	14,270	1	99.993
ID-IDplex	669	16	10,704	19	99.822
ID-PP16	662	14	9,268	4	99.957
ID-MiniFiler	1308	9	11,772	27	99.771
SGM-NGM	1436	11	15,796	4	99.975
ID-NGM	1449	11	15,939	3	99.981
ProPlus-NGM	1427	10	14,270	4	99.972
SGM-ESI	1436	1 mill	ion com	narisona	99.968
ProPlus-ESX	1427	- 1 11111		parisona	9 9.970
ESI-ESX	1455	>1100 dif	ferences	observ	<mark>ed</mark> 99.939
ESI-ESSplex	1445	00.00			<mark>99.875</mark>
ESX-ESSplex	1445	~99.9		raance	<mark>99.870</mark>
ESI-NGMSElect	715	(man	v correcte	d now)	99.860
ESX-NGMSElect	715		12,100		99.942
ESS-NGMSElect	663	17	11,271	17	99.849
		TOTAL	240,156	186	99.923

Kits (except Identifiler) were kindly provided by **Applied Biosystems, Promega, and Qiagen** for concordance testing performed at NIST

Extra (Degenerate) Primers Added with NGM SElect NGM SElect

NGM (original)

<u>11,11</u>

D2S441

9.1 allele missing in 7 Asians

D22S1045

15 allele missing in 4 samples

Amelogenin

X allele missing in 3 samples





and NGM'





15,17

X,Y



NIST Standard Sample Sets

• U.S. Population Samples (663 samples)

- Previously studied with Identifiler, MiniFiler, Yfiler, PP16, PP
 ESX/ESI 17, NGM, miniSTRs, and 23plex (>200,000 allele calls)
- 260 African Americans, 260 Caucasians, 140 Hispanics, and 3 Asians
- U.S. Father/Son pairs (800 samples)
 - Previously studied with Identifiler, MiniFiler, Yfiler, PP ESX/ESI 17, NGM, 23plex
 - ~100 fathers/100 sons for each group: African Americans, Caucasians, Hispanics, and Asians
- NIST SRM 2391b PCR DNA Profiling Standard (12 samples)
 - Components 1-10 (includes 9947A and 9948): well characterized
 - ABI 007 and K562

>1450 total samples

http://www.cstl.nist.gov/biotech/strbase/NISTpop.htm



D 004470	Allele	Promega	ABI	Repeat Structure	Reference
<u>D851179</u>	(Repeat #)	PowerPlex 16	Identifiler	[TCTR] _n	
	6	199 bp	119 bp	Not published	STRBase
	7	203 bp	123 br	[TCTA] ₇	Griffiths <i>et al.</i> (1998)
Known Alleles	8	207 bp	127 op	[TCTA] ₈	Barber and Parkin (1996)
	9	211 bp	131 bp	[TCTA] ₉	Barber and Parkin (1996)
	10	215 bp	135 bp	[TCTA] ₁₀	Barber and Parkin (1996)
	10.1	216 bp	136 bp	Not published	STRBase
	10.2	217 ор	137 bp	Not published	STRBase
	11	219 bp	139 bp	[TCTA] ₁₁	Barber and Parkin (1996)
	12	223 bp	143 bp	[TCTA] ₁₂	Barber and Parkin (1996)
	12.1	224 bp	144 bp	Not published	STRBase
Many alleles	12 2	225 bp	145 bp	Not published	STRBase
marry ancies	12.3	226 bp	146 bp	Not published	STRBase
sequences	13 (a)	227 bp	147 bp		Barber and Parkin (1996)
are not	13 (b)	227 bp	147 bp	[TCTA] ₂ [TCTG] ₁ [TCTA] ₁₀	Kline <i>et al.</i> (2010)
	13 (c)	227 bp	147 bp	[TCTA] ₁ [TCTG] ₁ TGTA[TCTA] ₁₀	Kline <i>et al.</i> (2010)
known	13 (d)	227 bp	147 bp	[TCTA] ₁₃	Kline <i>et al.</i> (2010)
	13.1	228 bp	148 bp	Not published	STRBase
	13.2	229 bp	149 bp	Not published	STRBase
	13.3	230 bp	150 bp	Not published	STRBase
	14	231 bp	151 bp	[TCTA] ₂ [TCTG] ₁ [TCTA] ₁₁	Barber and Parkin (1996)
	14.1	232 bp	152 bp	Not published	STRBase
We just set the	14.2	233 bp	153 bp	Not published	SIRBase
we just set the	15	235 bp	155 bp		Barber and Parkin (1996)
new world record	15.1	236 bp	156 bp	Not published	SIRBase
for the largest D8	15.2	237 bp	157 bp	Not published	SIRBase
for the largest be	15.3	238 00	158 bp	Not published	SIRBase
allele ("24 ")	16	239 bp	159 bp	$[ICIA]_2[ICIG]_1[ICIA]_{13}$	Barber and Parkin (1996)
	16.1	240 bp	160 bp	Not published	SIRBase
	17	243 bp	163 bp	$[ICIA]_2[ICIG]_2[ICIA]_{13}$	Barber and Parkin (1996)
	17.1	244 bp	164 bp	Not published	SIRBase
	17.2	245 DP			SIKBASE
	10	247 DP	107 D 171 bp	$[I \cup IA]_2[I \cup IG]_1[I \cup IA]_{15}$	Darber and Parkin (1996)
	19			$[I \cup [A]_2[I \cup [G]_2[I \cup [A]_{15}]$	
	20	255 bp	175 bp	ivot publisnea	STRBase

D8S1179 12, "24"



Allele 12 : [TCTA]₂ TCTG [TCTA]₉

Allele "24" : [TCTA]₂ TCTG [TCTA]₉ duplication of the 48 bases 10 bases downstream of the repeat

Result with This Large D8S1179 Allele Using European STR Kits

NGM SElect

PP ESX 17

PP ESI 17



False D21S11 tri-allele

False FGA tri-allele

Reverse primer internal to duplicated flanking region

ABI 3500 Genetic Analyzer

Status Update on Open Letter to Applied Biosystems Open Letter to Applied Biosystems on Concerns with ABI 3500

- 3/14/11 emailed ~900 forensic DNA scientists (SWGDAM, forens-dna, ENFSI, EDNAP) inviting them to sign onto a letter that will be sent to Applied Biosystems expressing concern with ABI 3500
- Very positive response with 101 who agreed to sign the letter
- Letter was sent March 31 to the president of ABI and scientists involved with the ABI 3500
- Community will be notified of ABI's response

A Sampling of Feedback I Received...

- People did not just sign the letter but many have an opinion about the issues or concern about ABI customer support (I have received >100 emails – often with some very strong thoughts)
- "I think that the AB3500 related issues most likely represent the beginning of a sea of problems, against which every independent lab must take arms. It is not up to the manufacturer of a machine to decide the basic procedures of a lab - it is up to the lab" (4/29/11)
- "I greatly appreciate your advocacy on behalf of our community. **Hopefully we will be heard**." (4/1/11)

Concerns Expressed in Open Letter

RFID tags

00 Series

- New .hid file structure requires new software
- Short shelf life of reagents would like to see data for expiration times

Hopefully a change will result...

A desire for greater communication with the community – the 3500 FAQ sheet is a good start but does not directly address all of the concerns raised

What was learned from ABI visit to NIST on May 11, 2011

- RFID over-ride is possible (their R&D lab has instrument that can use "expired" reagents)
- New software is required for 3500 .hid or .fsa files due to new file structure
- They do not have ANY data to support short shelf life of 3500 reagents
 - hard stops keep labs from having failures that lead to ABI having to replace arrays
- ABI 31xx instruments have a 4X signal reduction

Support to the Community

...Bringing traceability and technology to the scales of justice...

- Conduct interlaboratory studies
- Perform beta-testing of new human identity testing products
- We collaborate with other NIJ grantees
- We provide input to (or have aided):
 - Scientific Working Group on DNA Analysis Methods (SWGDAM)
 - Department of Defense Quality Assurance
 Oversight Committee for DNA Analysis
 - Virginia DFS Science Advisory Committee
 - American Prosecutor's Research Institute (APRI) DNA Forensics Program "Coursein-a-Box" for training lawyers
 - WTC Kinship and Data Analysis Panel (KADAP) and Hurricane Katrina efforts
 - NIJ Expert System Testbed (NEST) Project



Thank you for your attention

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Contact Information

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http://www.cstl.nist.gov/biotech/strbase

Our team publications and presentations are available at: http://www.cstl.nist.gov/biotech/strbase/NISTpub.htm